

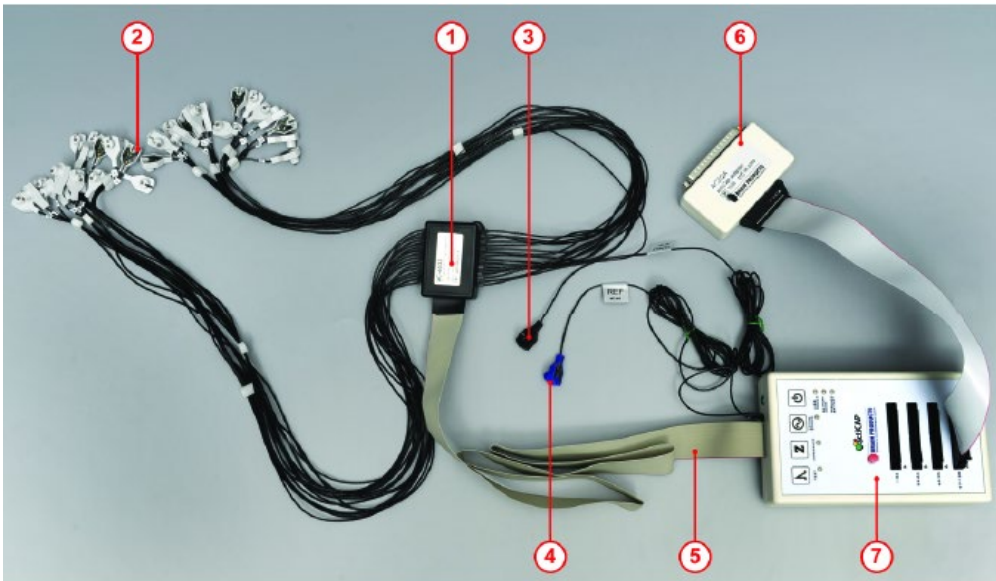
Disinfection and cleaning procedure EEG equipment

Caps and electrodes should be cleaned immediately after use, by following these steps:

Remove the electrode holder from the participant's chair and place it next to the sink in the white insert. Place the electrodes not directly in the sink, but **in the colander** in order to avoid damage to the electrode tips.

Make sure that the **splitter box and connection cable** (no 1 and 5 in figure) **are kept dry at all times!**

Do not pull the electrode wires at any point!



Remove the electrodes from the cap using a special *remover tool* (see pictures on the right). Hold the cap with one hand (it helps to put your thumb on the electrode) and bring the remover tool in between the ring and the electrode head. **Reach from the front of the electrode**, not from the tail!



Clean the electrodes (electrode tips and side opening of the electrode head) with a *toothbrush* using *lukewarm, running water*. If the electrodes have a tubular space, make sure to clean it with a *needle or tooth pick*! While doing this hold the electrode by the head, not the very tail of the plastic part and **not the wire!**

Clean the wires of the electrodes once a day by wiping them with a *baby towel*. Make sure all gel remains are removed from wires and electrodes!

Rinse the electrodes with *distilled water*. You can reuse the distilled water for all your participants within one day. Put the electrode wires on the splitter box hanger and return the splitter box hanger to the support, which is placed on the table in the experiment room.

Clean the (holes in the) cap with a *small dental brush* (see photo on the right, **do not** use this brush for cleaning the electrodes, as **this can cause damage!**). Put the brush through each hole and remove the gel on the brush with your fingers before pulling the brush back. Make sure that all gel remains are removed from the cap! Next, clean the syringes with warm water.



Disinfect the cap and the syringes (not the electrodes) with *Sekusept*¹:

Wear suitable protective clothing: gloves.

Produce the disinfection solution in the designated bucket: Mix **40 ml** of Sekusept with **960 ml** water.

Soak cap and syringes **in the solution for 15 minutes, not longer!**

Rinse with tap water.

The Sekusept solution is to be prepared daily. You can reuse the solution for all participants within one day.

Hang the cap to dry. Any residual dampness can dry off in the air. Using the hairdryer to speed up drying is not encouraged as the hot air can ruin the elasticity of the cap. If using the hair dryer can't be avoided due to consecutive measurements, use **medium** temperature and blow settings (e.g., not the highest).

Hang the used or wet towels to dry outside the lab on the dryers and put dried towels in the laundry basket.

DO NOT PUT WET TOWELS IN THE LAUNDRY BASKET!

Remove gel remains on table, chair, and equipment.



Put the needles in the separate **carton box for needle disposal**, not in the bin.

The last person scheduled in the lab will be held responsible for its status. If caps, electrodes or other parts of the lab were insufficiently cleaned you will be contacted by the lab coordinator.

Sekusept: In case of contact with eyes, rinse immediately with plenty of water and seek medical advice. If in contact with skin immediately wash off with copious quantities of water. Remove stained/soaked clothes immediately. Keep out of reach of children. Store in original container below 25 degree. Do not mix disinfectants with other products. Do not empty concentrated Sekusept Plus in the sink.



¹ Sekusept can be used for the disinfection and cleaning of thermolabile and thermostable medical instruments, endoscopes and electrodes. Effective against bacteria (TBC), viruses (HIV, HBV). 100 g contains agents: 25 Glucoprotamil.